of the pedicels is frequent in the genus and at least until other more convincing differences are found this plant of Friedrichsthal may be provisionally placed as

B. Kellermanii, forma **podocephala**, forma nov., formae typicae simillima differt capitulis graciliter pedicellatis; pedicellis 3-10(-20) mm. longis erectis vel adscendentibus.—Guatemala: Friedrichsthal (K., phot. and fragm. Gr.).

II. STUDIES IN THE BORAGINACEAE.

By I. M. Johnston

1. RESTORATION OF THE GENUS HACKELIA.

As currently taken the genus Lappula is composed of two sharply differentiated groups. It is here proposed that the perennial and biennial species with pyramidal gynobase be segregated to form the genus Hackelia, while the annual species with subulate gynobase be left to constitute the genus Lappula. The very important characters which separate these very distinct genera may be realized by a study of the following contrast.

LAPPULA. Annual; inflorescence abundantly bracteate; pedicels erect; gynobase subulate, 5–10 times as tall as broad, about equaling the nutlets; style surpassing the nutlets; nutlets narrowly attached all

along the well developed medial ventral keel.

HACKELIA. Biennial or perennial; inflorescence naked or rarely sparsely bracteate; pedicels recurved or deflexed in fruit; gynobase pyramidal, less tall than broad; style definitely surpassed by nutlets; nutlets attached by a large oblique submedial ovate or deltoid areola; ventral keel extending over only upper half of nutlet.

As usually taken Lappula has been an unnatural aggregate formed of two groups whose structures are so different that it seems improbable that the groups are immediately related. The species which I have referred to Hackelia do not find their nearest relations among the species of true Lappula, but rather among the species in section Coloboma of Eritrichium. Indeed so close and unmistakable are the relations between Hackelia and Eritrichium that with much justification the two genera might be merged. On the other hand Hackelia has been referred to Lappula only because the species in both genera have glochidiate bristles on the dorsal rim of the nutlets, and despite the fact that the species of the two groups differ markedly in habit and in a number of fundamental characters. As both Lappula and

Hackelia have in all probability evolved from the Cynoglosseae where glochidiate bristles are common as carpel appendages, it must be evident that the presence of barbed appendages is only of general phylogenetic significance and not a safe criterion upon which to judge immediate relationships.

Hackelia and Lappula differ in the manner by which the nutlets are attached to the gynobase. Among the borages this correctly has been considered of fundamental importance. In the characters of gynobase and nutlets Hackelia is almost exactly matched by developments in Eritrichium § Coloboma, for many species of that section approximate upon a reduced scale the characteristic habit of Hackelia, while furthermore some species, such as E. strictum Decne., E. pectinatum (Pall.) A.DC., etc., have the toothing on the dorsal margining of the nutlet tipped by subulate barbed prolongations.

Although approaching one another closely Hackelia and Eritrichium seem readily distinguished by habit, the former being rankgrowing green biennials or perennials with broad thin leaves and stems 2-10 dm. high, whereas Eritrichium contains low, canescent or silvery, strongly rooted, caespitose plants with small firm leaves and stems 1-20 cm. high. The dorsal margining of the nutlets in Hackelia is usually broken up into flattened subulate glochidiate appendages, but in Eritrichium the margin is entire or merely dentate and usually without glochids. The only invariably diagnostic character by which the two genera can be separated is that found in the direction of the fruiting pedicels; in Hackelia the pedicels are recurved or reflexed in fruit whereas in Eritrichium they are always erect or nearly so. Hackelia centers in western North America and has outlying species in the Andes and Himalayas, and in central Europe, eastern United States, and Mexico. Eritrichium centers in Asia, but has an arctic series of species occurring at high latitudes or altitudes in Europe and North America.

The genus Hackelia was published for Opiz by Berchtold in the latter's "Oekonomisch-technische Flora Böhmens." The genus was separated from Echinospermum for reasons unstated, but evidently not because of the characters stressed now in resurrecting it, for although H. deflexa was the only species fully described under the genus, there were eight of Ledebour's species, all true Lappulas, referred to it in a footnote. The characteristic areola and attachment of the nutlets are described in the lengthy specific diagnosis of H. deflexa, but in the generic description the style and the attachment of the nutlets are described but vaguely, and in similar terms as in the

generic description of *Echinospermum* a few pages further on. *Hackelia deflexa* (Wahl.) Opiz, being the first species given under *Hackelia* and the only one treated with any detail, it is taken as constituting the type of the genus as here redefined. The species referable to *Hackelia* are as follows:—

Hackelia deflexa (Wahl.) Opiz in Bercht. Fl. Böhm. ii, pt. 2, 147 (1839). Myosotis deflexa Wahl. Vet. Acad. Handl. Stockholm, xxxi. 113, t. 4 (1810). Echinospermum deflexum Lehm. Asperif. i. 120 (1818). Rochelia deflexa R. & S. Syst. iv. 109 (1819). Cynoglossum deflexum Roth, Enum. i. 589 (1827). Lappula deflexa Greene, Pittonia ii. 182 (1891). Echinospermum deflexum, var. americanum Gray, Proc. Am. Acad. xvii. 224 (1882). Lappula deflexa, var. americana Greene, l. c. 183. L. americana Rydb. Bull. Torr. Cl. xxiv. 294 (1897).

H. virginiana (L.), comb. nov. Myosotis virginiana L. Sp. Pl. 131 (1753). Echinospermum virginicum Lehm. Asperif. i. 117 (1818). Rochelia virginiana R. & S. Syst. iv. 108 (1819). Lappula virginiana Greene, Pittonia ii. 182 (1891). Cynoglossum Morisoni A.DC.

Prodr. x. 155 (1846).

H. pinetorum (Greene), comb. nov. Echinospermum pinetorum Greene in Gray, Proc. Am. Acad. xvii. 224 (1882). Lappula pinetorum Greene, Pittonia ii. 182 (1891). L. pustulata Macbr. Contr. Gray Herb. xlviii. 39 (1916). L. heliocarpa Brand in Fedde, Repert.

xviii. 310 (1922).

H. Roylei (Wall.), comb. nov. Cynoglossum Roylei Wall. in Don, Gen. Syst. iv. 356 (1838). C. laxum Don, l. c. Lappula laxa Macbr. Proc. Am. Acad. li. 543 (1916). C. uncinatum Royle, acc. to Benth. in Royle, Ill. 305 (1839). C. glochidiatum Wall. acc. to Benth. in Royle, l. c. 306. Echinospermum glochidiatum A.DC. Prodr. x. 136 (1846). Paracaryum glochidiatum Benth. in Hook Fl. Brit. Ind. iv. 161 (1883). L. glochidiata Brand in Fedde, Repert. xiv. 146 (1915).

H. macrophylla (Brand), comb. nov. Lappula macrophylla Brand in Fedde, Repert. xiv. 147 (1915). Cynoglossum uncinatum, var. laxiflora Benth. in Royle, Ill. 305 (1839). Echinospermum glochidiatum, var. laxiflorum A.DC. Prodr. x. 136 (1846).—From their descriptions this and the next species seem scarcely distinct from the preceding one.

H. Dielsii (Brand), comb. nov. Lappula Dielsii Brand in Fedde,

Repert. xiv. 147 (1915).

H. revoluta (R. & P.), comb. nov. Cynoglossum revolutum R. & P. Fl. Peruv. ii. 6 (1799). Lappula revoluta Brand in Fedde, Repert.

xiv. 148 (1915). C. ovatifolium Griesb. Götting. Abhandl. xxiv. 271 (1879). L. revoluta, f. ovatifolia Brand, l. c. C. parviflorum Krause, Engler Bot. Jahrb. xxxvii. 634 (1906). C. Fiebrigii Krause, l. c. L. revoluta, f. Fiebrigii Brand, l. c. C. andicolum Krause, l. c. 635.

H. costaricensis (Brand), comb. nov. Lappula costaricensis Brand in Fedde, Repert. xviii. 310 (1922). L. guatemalensis Brand,

l. c. 311.

H. mexicana (Schl. & Cham.), comb. nov. Cynoglossum mexicanum Schl. & Cham. Linnaea v. 114 (1830). Echinospermum mexicanum Hemsl. Biol. Cent.-Am. Bot. ii. 377 (1882). Lappula mexicanum Greene, Pittonia ii. 182 (1891).—This and the preceding are closely related to H. revoluta and perhaps are not specifically distinct.

H. leptophylla (Rydb.), comb. nov. Lappula leptophylla Rydb. Mem. N. Y. Bot. Gard. i. 329 (1900). (?) L. scaberrima Piper, Bull. Torr. Cl. xxix. 545 (1902). (?) L. angustata Rydb. Bull. Torr. Cl. xxxi. 636 (1904). L. Besseyi Rydb. l. c. 636. (?) L. grisea Woot. & Standl. Contr. U. S. Nat. Herb. xvi. 164 (1913). L. floribunda of Piper (Bull. Torr. Cl. xxix. 537. 1902.) and other recent authors.

H. hispida (Gray), comb. nov. Echinospermum diffusum, var. hispidum Gray, Proc. Am. Acad. xvii. 225 (1882). E. hispidum Gray, l. c. xx. 259 (1884). L. hispida Greene, Pittonia ii. 182 (1891).

H. ciliata (Dougl.), comb. nov. Cynoglossum ciliatum Dougl. in Lehm. Pug. ii. 24 (1830). Echinospermum ciliatum Gray, Proc. Am. Acad. xvii. 225 (1882). Lappula ciliata Greene, Pittonia ii. 182 (1891).

H. setosa (Piper), comb. nov. Lappula setosa Piper, Bull. Torr. Cl. xxix. 544 (1902).

H. cinerea (Piper), comb. nov. Lappula cinerea Piper, Bull. Torr. Cl. xxix. 544 (1902).

H. ursina (Greene), comb. nov. Echinospermum ursinum Greene in Gray, Proc. Am. Acad. xvii. 224 (1882). Lappula ursina Greene, Pittonia ii. 182 (1891).

H. hirsuta (Woot. & Standl.), comb. nov. Lappula hirsuta Woot.

& Standl. Contr. U. S. Nat. Herb. xvi. 164 (1913).

H. gracilenta (Eastw.), comb. nov. Lappula gracilenta Eastw. Bull. Torr. Cl. xxix. 523 (1902).

H. floribunda (Lehm.), comb. nov. Echinospermum floribundum Lehm. Pug. ii. 24 (1830). Lappula floribunda Greene, Pittonia ii. 182 (1891). Rochelia patens Nutt. Jour. Acad. Philad. vii. 44 (1834). E. subdecumbens Parry, Proc. Davenport Acad. i. 148 (1876). L.

subdecumbens Nels., Manual Rocky Mt. Bot. 412 (1909). (?) L. leucantha Greene, Leaflets i. 152 (1905). L. Jessicae McGreg. Bull. Torr. Cl. xxxvii. 262 (1910). L. diffusa of Piper and other recent authors.—The plant referred here has always been associated with Lehmann's Echinospermum diffusum despite the fact that it is perfectly portrayed in Hooker's, Fl. Bor. Am. ii. 84, t. 164 (1838), plate of E. floribundum, a plate apparently drawn from an isotype if not the actual type of that latter species. It is to be particularly noted that the plant, the Lappula diffusa of recent authors, which I refer to Lehmann's Echinospermum floribundum agrees with the pictured plant in perennial duration and in size of corolla, developments not present in H. leptophylla, the plant usually referred to Lehmann's species. In spite of the note by Nelson and Macbride, Bot. Gaz. lxi. 42 (1916), the account given by Parry, l. c., and the isotype preserved in the Gray Herbarium both clearly show that Gray, Proc. Am. Acad. xvii. 225 (1882), and Piper, Bull. Torr. Cl. xxix. 539 (1902). were correct in referring Echinospermum subdecumbens to the present species. The use of Parry's name in the Rocky Mountain Manual is incorrect! The plant is definitely blue-flowered, as the lack of mention of color in Parry's observations would suggest.

H. Eastwoodae, nom. nov. Lappula micrantha Eastw. Bull. Torr. Cl. xxx. 497 (1903); not H. micrantha (Ledeb.) Opiz.—Related to H. floribunda, but differing in small flowers and in having a few glochidiate prickles on the back of the nutlets. Perhaps only the Californian variety of that species.

H. bella (Macbr.), comb. nov. Lappula bella Macbr. Contr. Gray Herb. xlviii. 39 (1916). (?) L. Rattanii Brand in Fedde, Repert. xviii. 311 (1922).—Differing from the closely related H. velutina in possessing white corollas, and in having the back of the nutlets unarmed.

H. nervosa (Kell.), comb. nov. Echinospermum nervosum Kell. Proc. Calif. Acad. ii. 146, f. 42 (1862). Lappula nervosa Greene, Pittonia ii. 182 (1891).

H. velutina (Piper), comb. nov. Lappula velutina Piper, Bull. Torr. Cl. xxix. 546 (1902).—Perhaps only a good variety of the last. The Hackelias of the Yosemite Region seem best referred here although they vary as to pubescence and have small corollas with short tubes.

H. californica (Gray), comb. nov. Echinospermum californicum Gray, Proc. Am. Acad. xvii. 225 (1882). Lappula californica Piper, Bull. Torr. Cl. xxix. 546 (1902).

VH. arida (Piper), comb. nov. Lappula arida Piper, Bull. Torr. Cl. xxviii. 44 (1901). L. Cottoni Piper, l. c. xxix. 549 (1902).

J. H. arida, var. Cusickii (Piper), comb. nov. Lappula Cusickii Piper, Bull. Torr. Cl. xxix. 542 (1902). L. arida, var. Cusickii Nels. & Macbr. Bot. Gaz. lxi. 41 (1916). L. saxatilis Piper, l. c. 541.

H. diffusa (Lehm.), comb. nov. Echinospermum diffusum Lehm. Pug. ii. 23 (1830). Lappula diffusa Greene, Pittonia ii. 182 (1891). L. Hendersoni Piper, Bull. Torr. Cl. xxix. 539 (1902). (?) L. trachyphylla Piper, l. c. 540. L. subdecumbens of Nels. Man. Rocky Mt. Bot. 412 (1909), as to description only.—The plant concerned here agrees with Lehmann's description in height of growth, pubescence, size and color of flowers, and in the arming of the nutlets. It fits the description far better than the other plant, here called H. floribunda, which has borne Lehmann's specific name in the past. In fact the present plant diverges from Lehmann's description only by having usually acutish rather than obtusish cauline leaves.

H. diffusa var. caerulescens (Rydb.), comb. nov. Lappula caerulescens Rydb. Mem. N. Y. Bot. Gard. i. 328 (1900). L. subdecumbens caerulescens Garrett, Fl. Wasatch Reg. 78 (1911).

2. The Genus Antiphytum.

In the literature concerned with the Boraginaceae the species of Antiphytum have been repeatedly confused with the species now referred to Plagiobothrys, Cryptantha, and allied genera, despite the fact that the former genus appears to belong not to the Eritrichieae, but rather to the Lithospermeae. This confusion has resulted from the attention being too closely centered upon similarities in nutlets, similarities which caused Gray, Proc. Am. Acad. xx. 265 (1885), to refer the Mexican species of Antiphytum to Krynitzkia, and which gave Macbride, Contr. Gray Herb. xlviii. 41 (1916), his reasons for saying that Amblynotopsis, here referred to Antiphytum, is "intermediate between the genera Allocarya and Plagiobothrys." The genus Antiphytum, however, differs from Plagiobothrys in habit, color of flowers, and particularly in the geminate stigmas. The stigmas of Antiphytum definitely remove it from the Eritrichieae in which Plagiobothrys and its other supposed allies are found.

The genus Antiphytum was first described in Meisner's Genera, i. 280 (1836-43). The original description, about fifty words in length, applies well to the genus as it has been taken in the past. In the Commentary accompanying his Genera, ii. 188 (1836-43), Meisner gives the bibliography of the genus as follows:—

"Antiphytum. DC. Mss. in Moç. ic. fl. Mex. ined. (4 sp.)—Anchusa oppositifolia, H.B.K., nov. gen. 3. p. 91. t. 200. A. cruciata et stoechadifolia, Cham. in Linnaea, 4. p. 438. et A. Mexicana, DC. ap. Moç.

l. c.—Genus jam foliis oppositis (unde nomen) insigne."

Mociño's flora referred to was unpublished until 1874, or over thirty years after the appearance of Meisner's Genera. Antiphytum mexicana DC., published in the Prodromus, x. 121, in 1846, is Heliotropium calcicola Fernald, Proc. Am. Acad. xliii. 62 (1907). Although the only Antiphytum published in Mociño posthumous work, A. mexicana can scarcely be considered as the type of the genus since it remained so long unpublished and particularly since having typical Heliotropium fruit, it is not at all described in the original generic diagnosis. Anchusa oppositifolia HBK. is an Allocarva. species being the first published species mentioned under Antiphytum and the only one with a cited illustration, might be considered the type of Antiphytum and that name treated as an older synonym of Greene's genus Allocarya, Pittonia i. 12 (1887). Usage, however, has restricted the name Antiphytum to the genus including Chamisso's Anchusa cruciata and A. stoechadifolia. This usage is justifiable by the workings of the logical process of residues, the other species originally placed in Antiphytum having been referred to other genera and the name Antiphytum left to that group which had a majority representation in the original definition of the genus. It is also to be noted that with the exception that they are never herbs, Chamisso's species agree with every item in Meisner's diagnosis. On the other hand Anchusa oppositifolia is never suffrutescent, and does not have scorpioid cymes, nor ciliate-papillose faucal protuberances.

In 1916 Macbride, l. c., erected the genus Amblynotopsis for the Mexican species which I refer to Antiphytum. Although four of the five recognized and previously published species had been at one time or another referred to Antiphytum no contrast was made between the members of the newly proposed genus and the old restricted one. A careful study of this relationship now shows that the Mexican plants, forming the genus Amblynotopsis, are distinguished from the Brazilian ones, forming the true Antiphytum, only by having non-blue corollas and alternate upper leaves. These are scarcely generic differences. The Mexican Antiphytum heliotropioides besides agreeing with the Brazilian A. cruciatum in shrubby habit and loose strigose pubescence of similar encrusted hairs, also has opposite leaves. The chief difference between the two plants being that in the Mexican plant the leaves of the inflorescence are alternate, whereas in the Brazilian

species the leaves are opposite throughout the plant. Among the Mexican species A. floribundum has all its leaves alternate. Such species as A. peninsulare and A. nudicalces have several pairs of opposite leaves. Hence within the enlarged Antiphytum there are all the stages from an entirely opposite-leaved condition to a completely alternate-leaved one. Since leaf-position has various degrees of development among the Mexican species and since the difference separating the Mexican and Brazilian species is simply a matter of slight degree it seems inadvisable to attempt the use of leaf-position as a generic character. Flower-color is equally unsatisfactory as a generic character. The Brazilian species have bluish flowers. Among the Mexican species A. peninsulare has white flowers, whereas the remainder have yellow ones. If the color of corolla is to be exalted to generic importance there will be need of three instead of merely two genera.

The Brazilian species have flat gynobases and nutlets that are basally attached by a short stipe-like prolongation. Although most Mexican specimens have their nutlets directly attached to a more or less pyramidal gynobase by a large oblique submedial ventral scar, certain specimens (Palmer 443, 207) here referred to the polymorphous A. floribundum have nutlets with basal attachments through a short stipe to a flattened gynobase quite like that exhibited in Brazilian specimens. A synopsis and bibliography of the genus follows:-

ANTIPHYTUM A.DC. in Meisner, Genera i. 280; ii. 188 (1836-43). Thaumatocaryon Baill. Bull. Mens. Soc. Linn. Paris 839 (1890). Amblynotopsis Macbr. Contr. Gray Herb. xlviii. 41 (1916).

Leaves all opposite; corolla bluish; South American. Plant herbaceous; leaves 10-35 mm. broad; nutlets smooth, shiny; corolla

Pedicels elongating, becoming 5-10 mm. long; leaves alternate only in the inflorescence; corolla with definite

rosettes, alternate, or only lowermost opposite. Corolla subrotate, throat broad and open, tube practically undeveloped; appendages lacking, stamens ex-

Stems strictly erect, 3-8 dm. high; basal leaves few,

 Corolla salverform, tube cylindrical and well developed; appendages well developed; stamens included and hidden.

Corolla white; fruiting calyx 8 mm. long, sparsely

corolla white; fruiting calyx 8 mm. long, sparsely strigose-hispid; plant bushy, about 5 dm. tall....6. A. peninsulare.

Corolla yellow; fruiting calyx 2-5 mm. long, densely strigose; plant caespitose or with a prostrate shrubby caudex, 1-2 dm. high.

Plant with a loosely branched shrubby caudex; leaves not in basal rosettes, all cauline......8. A. nudicalces.

Plant densely caespitose; basal leaves in dense

Plant densely caespitose; basal leaves in dense rosettes.

Corolla about 5 mm. broad; style surpassing nut-

Corolla about 1 mm. broad; style not surpassing

1. Antiphytum tetraquetrum (Cham.) A.DC. Prodr. x. 122 (1846). Anchusa tetraquetra Cham. Linnaea viii. 113 (1833). Thaumatocaryon Hilarii Baill. Bull. Mens. Soc. Linn. Paris 839 (1890). Antiphytum Bornmülleri Pilger in Fedde, Repert. iii. 24 (1906). Antiphytum Bornmülleri, var. asperior Pilger I. c. 25.

2. A. CRUCIATUM (Cham.) A.DC. Prodr. x. 121 (1846). Anchusa cruciata Cham. Linnaea iv. 438 (1829). Anchusa stoechadifolia Cham. l. c. 439. Antiphytum staechadifolium A.DC. l. c. Myosotis Berroi Arech. Anal. Mus. Nac. Montevideo, ser. 2, i. 69, f. 5-6 (1911). —The lengthy descriptions given by Chamisso do not seem to contain any fundamental characters by which his two species can be distinguished, nor has any subsequent writer pointed out diagnostic characters.

3. A. HELIOTROPIOIDES A.DC. Prodr. x. 122 (1846). Eritrichium heliotropioides Torr. Bot. Mex. Bound. 140 (1859). Krynitzkia heliotropioides Gray, Proc. Am. Acad. xx. 265 (1885). Cryptantha heliotropoides Loes. in Fedde, Repert. xii. 243 (1913). Amblynotopsis heliotropioides Macbr. Contr. Gray Herb. xlviii. 41 (1916).

- 4. A FLORIBUNDUM (Torr.) Gray, Proc. Am. Acad. x. 55 (1875). Eritrichium floribundum Torr. Bot. Mex. Bound. 140 (1859). Krynitzkia floribunda Gray, l. c. xx. 265 (1885). Amblynotopsis floribunda Macbr. Contr. Gray Herb. xlviii. 41 (1916). Amblynotopsis durangensis Macbr. l. c. 42.—As here taken the species is extremely polymorphous and almost certainly capable of division, but at present the material is too meager to attempt satisfactory segregation.

5. A. paniculatum, nom. nov. Lithospermum linifolium Mart. & Gal. Bull. Acad. Belg. xi. 338 (1844); not Antiphytum linifolium A.DC. (1846).

√6. A. peninsulare (Rose), comb. nov. Krynitzkia peninsularis Rose, Contr. U. S. Nat. Herb. i. 85 (1890). Amblynotopsis peninsularis Macbr. Contr. Gray Herb. xlviii. 41 (1916).

- 7. A. caespitosum, sp. nov., mexicanum; caulibus erectis vel valde ascendentibus 8–16 cm. altis apicem versus pauce stricteque ramosis; foliis dense strigosis argyro-canescentibus 1–2 mm. latis acutiusculis, inferioribus anguste linearibus 2–5 cm. longis erectis rosulatis, caulinis 8–15 mm. longis; racemis paucis unilateralibus manifeste bracteatis 2–3 cm. longis; calyce 2.5–3 mm. longo 5-partito breve pedicellato; corolla flava, limbo 4–5 mm. lato plano, lobis rotundatis imbricatis extus pubescentibus, tubo 1.5–2 mm. longo ad apicem cum 5 appendiculis gibbosis instructo, staminibus inclusis cum filamentis antheris brevioribus; nuculis rugoso-tuberculatis 1.5–2 mm. longis, areolis amplis distincte sub medio locatis; gynobasi angusta pyramidali; stylo nuculis longiore; stigmatibus geminatis.—Mexico: Cerros near San Luis Tultitlanapa, Puebla, Purpus 2606 (Type, Gray Herb.). Sonnige Kalkhügel bei Comitan, Chiapas, Seler 3073. Huauclilla, Nochixtlan, Oaxaca, Conzatti & González 1222.
- 8. A. nudicalces, sp. nov., diffusum; A. caespitosum similans sed differt ramis numerosis ramosis e caudice suffruticoso laxe ramoso prostrato vel etiam paullo subterraneo orientibus et foliis majoribus 2-3 mm. latis omnibus caulinis.—Mexico: Sosola, alt. 7000 ft., Oaxaca, L. C. Smith 393 (TYPE, Gray Herb.). Although differing from A. caespitosum conspicuously in habit this species is identical with it in inflorescence, floral, and fruit characters and may prove to be worthy of no more than varietal recognition.

9. A. Parryi Wats. Proc. Am. Acad. xviii. 122 (1883). Krynitzkia Parryi Gray, Proc. Am. Acad. xx. 265 (1885). Amblynotopsis Parryi Macbr. Contr. Gray Herb. xlviii. 41 (1916).—Mexico: En route from San Luis Potosi to San Antonio, Texas, Parry (Type). Sosola, Oaxaca, L. C. Smith 394. Without locality, Coulter 1050 in part. The type is in advanced maturity and entirely lacks flowers. It is associated with the small-flowered Oaxacan plant only because of the remarkable similarity in the size and developments of all other parts.

3. Novelties and New Combinations in the Genus Cryptantha.

Cryptantha Abramsii, sp. nov., annua basem versus simplex supra sparse ascendenter ramosa 15–30 cm. alta strigosa; foliis linearibus vel lineari-filiformibus 1–3 cm. longis 1–1.5 mm. latis sessilibus acutiusculis basem versus hispidis, infimis oppositis; spicis solitariis vel geminatis 2.5–10 cm. longis conspicue bracteatis, bracteis linearibus vel lanceolatis; corolla evidenti 1.5–2 mm. lata; calycibus maturitate 3–4 mm. longis remotis non biseriatis strictis vel ascendentibus, lobis lanceolatis breviter hispidis calyce ½3–¼ brevioribus

abaxillaribus crassissimis et hispidissimis; nuculis 1-4 lanceolatis laevibus nitidis basi truncatis compressis 2-2.2 mm. longis, sulcis clausis basem versus furcatis; stylo alto ½-½ nuculis breviori; gynobasi subulata.—Caltfornia: San Pedro Hills near Malaga Cove, Los Angeles Co., March 14, 1903, Abrams 3139 (Type, Gray Herb.; Isotype, Univ. Calif. Herb.). Allied to Cryptantha leiocarpa (F. & M.) Greene, but differing from that species in its short style, erect habit, and fewer nutlets. It suggests the large-flowered forms of C. Clevelandi Greene, and possibly may be only a bracteate form of

the latter species. Further material is a great desideratum.

Cryptantha Brandegei, sp. nov., annua diffusa decumbens; caulibus gracilibus 1-4 dm. longis strigosis pustulatis vel levibus; foliis oblongo-lanceolatis vel linearibus obtusis 6-15 mm. longis 2-3(-4) mm. latis basem versus sparse hispidis concoloribus; spicis solitariis vel rare geminatis aliquid sparse bracteatis vetustis 4-8 cm. longis; calycibus maturitate congestis vel remotis 2-4 mm. longis strictis; lobis calycis linearibus costatis cum setis flavescentibus horrentissimis marginibus sparse strigosis abaxillaribus longissimis hispidissimis; corolla parva minus quam 1 mm. lata; nuculis 1-4 laevibus 1.5-2 mm. lanceolatis nitidis basi truncatis, sulcis clausis basem versus furcatis; stylo alto nuculis $\frac{1}{3} - \frac{1}{5}$ breviori; gynobasi subulata.—California: Santa Rosa Island, June 1888, T. S. Brandegee (TYPE, Gray Herb.; ISOTYPE, Univ. Calif. Herb.). This is an ally of Cryptantha leiocarpa (F. & M.) Greene, from which it differs in its southern island occurence, greater range in nutlet-number, and particularly in its shorter style. It suggests phases of the polymorphous C. Clevelandi Greene, but is readily told from that species by its bracteate inflorescence and slightly longer style. Cryptantha Brandegei is to be distinguished from C. Abramsii by its smaller corolla and diffuse spreading habit.

Cryptantha albida (HBK.), comb. nov. Myosotis albida HBK. Nov. Gen. et Sp. iii. 91 (Aug. 1818). Lithospermum ramosum Lehm. Asperif. ii. 328 (Nov. or Dec. 1818). Eritrichium ramosum A.DC. Prodr. x. 132 (1846). Krynitzkia ramosa Gray, Proc. Am. Acad. xx. 274 (1885). Cryptanthe ramosa Greene, Pittonia i. 115 (1887). Eritrichium hispidum Buckley, Proc. Acad. Philad. 1861, 462 (1861). (?) Krynitzkia mexicana Brandg. Zoe v. 182 (1904).—Information kindly supplied me by Dr. J. H. Barnhart of the New York Botanical Garden has given the reason for reviving the long neglected Myosotis albida HBK. This name was published in the ninth part of the Nova Genera which, according to Dr. Barnhart, appeared probably

late in August 1818 since it was noted under the date of October 3, 1818 in the Bibliographie de la France, a work in which the appearance of books was usually announced about six weeks after their actual publication. The first part of Lehmann's Asperifoliae appeared before the ninth part of the Nova Genera and its priority was recognized by Kunth, Flora i. 601 (1818) and Nov. Gen. et Sp. iii. 451 (1820). When the first part of the Asperifoliae was reviewed in Flora, i. 501, under the date October 30, 1818, it was accompanied by the statement "Der zweyte Theil ist unter der Presse" which would seem to indicate that the second part of Lehmann's work did not appear for at least two months after the ninth part of the Nova Genera. The second part of the Asperifoliae may have appeared in November or December but at present there seems no way of telling whether it actually did appear before the end of 1818, the year given on the title page.

Cryptantha falcata (Hieron.), comb. nov. Eritrichium falcatum

Hieron. Bol. Acad. Córdoba iv. pt. 1, 64 (1882).

Cryptantha patagonica (Speg.), comb. nov. Amsinckia patagonica

Speg. Anal. Soc. Cient. Argent. liii. 137 (1902).

Cryptantha Spegazzinii, nom. nov. Amsinckia angustifolia, var. microcarpa Speg. Anal. Soc. Cient. Argent. liii. 136 (1902).—The nutlets described by Spegazzini are evidently not those of an Amsinckia. I am associating with this name a specimen from near General Roco, Rio Negro, Fischer 131, although the plant has the corolla and calyx subequal and not "corollae... calyce duplo longioris."

Cryptantha granulosa (R. & P.), comb. nov. Myosotis granulosa R. & P. Fl. Peruv. ii. 5 (1799).

Cryptantha corymbosa (R. & P.), comb. nov. Myosotis corymbosa R. & P. Fl. Peruv. ii. 5 (1799).

In 1887 Greene, Pittonia i. 58-60, proposed the genus Eremocarya and reëstablished Torrey's Piptocalyx. Since that time the two genera have received almost universal acceptance despite the fact that they appear to lack fundamental characters and much resemble members of the genus Cryptantha. The nutlets found in Eremocarya and Piptocalyx are indistinguishable from those of Cryptantha, being of similar shape, possessing similar markings, and having a very similar groove. The gynobase also is much the same in all three genera. In fact, Eremocarya and Piptocalyx seem merely well marked species of Cryptantha and are consequently referred to Cryptantha where their species can be disposed of as follows:—

Cryptantha § Piptocalyx. Piptocalyx Torr. in Wats. Bot. King Exped. 240 (1871); not Oliver (1870). Krynitzkia Subsect. Piptocalyx Gray, Proc. Am. Acad. xx. 275 (1885). Krynitzkia Sect. Piptocalyx Greene, Bull. Calif. Acad. i. 206 (1885). Greeneocharis Gürke & Harms in E. & P. Nat. Pflanzenf., Gesamtreg. 462 (1899). Wheelerella Grant, Bull. So. Calif. Acad. v. 28 (1906).—This section of Cryptantha is characterized by its peculiar circumscissile calyx which is tubular to above the middle and has its lobes practically unribbed. A short distance below the sinuses the calyx-tube suddenly changes, at the line of dehiscence, from firm siliceous-hvaline to herbaceous. In the characters of its calyx the plants much resemble certain species of Plagiobothrys. Among the suggested generic characters of Piptocalyx, Greene especially stressed its possession of persistent pedicels although that development is present in such Cryptanthas as C. albida, C. racemosa, C. holoptera, and C. pterocarya. The character most emphasized by Greene, however, was the dichotomy of Piptocalyx. Unfortunately this character also fails since unmistakable and very similar dichotomy occurs in such species as Cryptantha recurvata and C. micromeres. The only distinctive character possessed by Piptocalyx is its circumscissile calyx. This development, however, is both present and absent in the closely related genus Plagiobothrys and there seems no particular reason why in the present case the character should be considered of generic value. The following two species are recognized.

Cryptantha circumscissa (H. & A.), comb. nov. Lithospermum circumscissum H. & A. Bot. Beech. 370 (1840). Piptocalyx circumscissus Torr. in Wats. Bot. King. Exped. 240. (1871). Eritrichium circumscissum Gray, Proc. Am. Acad. x. 58 (1874). Krynitzkia circumscissa Gray, l. c. xx. 275 (1885). Wheelerella circumscissa Grant, Bull. So. Calif. Acad. v. 28 (1906). Greeneocharis circumscissa Rydb. Bull. Torr. Cl. xxxvi. 677 (1909). Cryptanthe depressa Nels. Bot. Gaz. xxxiv. 29 (1902).

Cryptantha dichotoma (Greene), comb. nov. Krynitzkia dichotoma Greene, Bull. Calif. Acad. i. 206 (1885). Piptocalyx dichotomus Greene, Pittonia i. 60 (1887). Wheelerella dichotoma Grant. Bull. So. Calif. Acad. v. 28 (1906). Greeneocharis dichotoma Macbr. Proc. Am. Acad. li. 546 (1916). G. circumscissa, var. hispida Macbr. l. c.—Macbride's variety of G. circumscissa is evidently a reduced montane form of C. dichotoma which simulates C. circumscissa in gross aspect. It makes it impossible to use robustness as a distinguishing character between C. dichotoma and C. circumscissa, and leaves pubescence as the only differentiating character.

According to Greene, Pittonia i. 56 (1887), "Eremocarya is most excellently marked in a three-fold way by its racemes" which are biserial and very dense, conspicuously bracteate, and repeatedly dichotomous. Neither singly nor in combination do these characters distinguish Eremocarya from Cryptantha. Almost every species of Cryptantha has its flowers somewhat biserial. In Cryptantha Grayi, C. albida, C. pusilla, C. maritima, etc., particularly dense biserial racemes may be found. Dichotomy is also frequently present in Cryptantha and is quite unmistakable in C. albida. Bracteate racemes are well developed in C. maritima, C. leiocarpa, C. albida, etc. Also emphasized by Greene was the dye-secreting tissue of Eremocarya. Following him most recent authors have dignified that development by treating it as the crucial generic character. In Plagiobothrys, even as limited by Greene, there are species with dye-secreting tissue and those without. This example would give precedent for including dye-secreting and non-dye-secreting species within the same genus, even were there no recognized case of dye-secretion among the indubitable species of Cryptantha. Dye-secretions in the roots are not uncommon in Cryptantha and in the Gray Herbarium are found present in specimens of such distinct species as C. Fendleri (Osterhout 3425, Patterson 112, Baker 780) and C. muricata (Parish 929). During 1921 I collected on the islands of the Gulf of California a yet unpublished variety of C. Grayi which has its roots as heavily charged with purple dye as do the most characteristic specimens of Eremocarya. In addition to the above characters, which are evidently insufficient to justify generic segregation, Greene gave Eremocarya as having "a persistent open calyx and an enlarged persistent style." The persistent open calyx of Eremocarya is well matched in C. holoptera and in C. albida, while in what Greene calls an "enlarged persistent style" Eremocarya is indistinguishable from the several species allied to true C. muricata. A careful study of Eremocarya has failed to reveal characters other than those unsatisfactory ones enumerated by its author and I am consequently forced to the conviction that Greene's genus is unworthy of recognition even as a section. Accordingly the following species and variety are referred to Cryptantha where they fit naturally into the same group of species as C. Grayi and and C. angustifolia.

Cryptantha micrantha (Torr.), comb. nov. Eritrichium micranthum Torr. Bot. Mex. Bound. 141 (1859). Krynitzkia micrantha Gray, Proc. Am. Acad. xx. 275 (1885). Eremocarya micrantha Greene, Pittonia i. 59 (1887). Eremocarya muricata Rydb. Bull. Torr. Cl. xxxvi. 677 (1909).

Cryptantha micrantha, var. lepida (Gray), comb. nov. Eritrichium micranthum, var. lepidum Gray, Synop. Fl. N. A. ii. pt. 1, 193 (1878). Krynitzkia micrantha, var. lepida Gray, Proc. Am. Acad. xx. 275 (1885). Eremocarya lepida Greene, Pittonia i. 59 (1887). Eremocarya micrantha, var. lepida Macbr. Proc. Am. Acad. li. 545 (1916).

4. A Synopsis and Redefinition of the Genus Plagiobothrys.

In 1835 the name *Plagiobothrys* was originally used by Fischer and Meyer for what then appeared to be a monotypic Chilean genus. The first species, *P. fulvus*, was separated from *Eritrichium* because of the peculiar annular scar on its nutlets. In 1874 Gray, Proc. Am. Acad. x. 57, reduced *Plagiobothrys* to a section under *Eritrichium* and placed in the section besides the original species five others which lacked annular scars on the nutlets. *Plagiobothrys* was reëstablished by Gray, Proc. Am. Acad. xx. 281, in 1885 when he amplified it to include fourteen species, five of which were placed in a newly erected section, and nine of which were put in his section *Genuini*, a group coëxtensive with his *Eritrichium* § *Plagiobothrys* of 1874.

Gray, Proc. Am. Acad. xi. 89, founded the genus Echidiocarya in 1876, and at that time included in it only the anomalous E. arizonica (P. Pringlei Greene). The character for the genus was found in the long-stiped nutlets. In 1877, Proc. Am. Acad. xii. 163, the genus was enlarged so as to include the newly described and obviously related E. californica. A third member of the group was added in 1883, Proc. Am. Acad. xix. 90, when Gray described P. ursinus and noted that, "The comparatively recent discovery of the preceding species [P. ursinus] of this section has made it clear that both of them should fall into Plagiobothrys, . . . " As a result of the transfer Echidiocarva was reduced to its original species and characterized by its "conspicuously stipitate" nutlets. In 1887 Greene, Pittonia i. 9 & 21, argued the artificiality of this latter concept and transferred to Plagiobothrys the remaining and type species of Echidiocarya saying that it had "every aspect and every character of Plagiobothrys, except that there is a stipe between the scar, or point of attachment to the gynobase, and the body of the nutlet." Greene's disposal of Echidiocarya has remained unchallenged.

Anyone who will study Gray's Echidiocarya arizonica, E. californica, and Plagiobothrys ursinus can not help appreciating the close relations between those species and the naturalness of Echidiocarya in its broadest sense, for the species agree not only in gross aspect, but in

scores of important and unimportant minute details as well. The fruit, though varying somewhat in the development of the stipe-like base, is similar in form and marking. Furthermore there is a strong similarity among the three species as regards calyx and corolla. None of the species have obvious relations within Plagiobothrys as that genus is currently taken. The three are sharply set off from it by their lack of a conspicuous caruncular scar on the nutlets and by their bearing the scar on a stipe projected above the ventral keel and not in a broad shallow transverse groove below the level of the keel. fact Echidiocarya is not a part of Plagiobothrys, as that genus is usually taken, but belongs rather to the group which Greene named Allocarya. This latter relation is indicated by the occurrence of lower opposite leaves in all three species. Significant also is the fact that the habit of Echidiocarya is approached by various species of Allocarya, and its nutlet-form simulated by the fruit of Allocarya australasica. species of Echidiocarya seem clearly congeneric with those of Allocarya. and since Gray's generic name is about ten years older than that of Greene the concept commonly called Allocarya will have to be called Echidiocarya if it be actually worthy of generic recognition.

The genus Sonnea was proposed by Greene, Pittonia i. 22, in 1887. It consists of two quite distinct and apparently not immediately related elements. The typical section of Sonnea contains the species which Gray called Plagiobothrys glomeratus and P. hispidus, and placed in the specially formed Plagiobothrys § Hypsoula. This group differs from all others of Plagiobothrys in its glomerate inflorescence, rough hispid pubescence, and supramedial caruncular scar. Although the group is a natural one and is fairly well marked, the form and structure of its nutlets reveal such unmistakable relations with Plagiobothrys that it seems best to consider it a member of that genus as most recent authors have been content to do. The fragile nature of the caruncle, particularly emphasized by Greene, is not positive nor capable of exact definition. It is not worthy of

particular note and is certainly not of generic value.

The second section of Sonnea is composed of three species falling under what Gray, Proc. Am. Acad. xx. 281, in 1885 indicated as Plagiobothrys * Ambigui. These species differ from those in genuine Plagiobothrys by having a coarse hispid pubescence and nutlets which superficially closely simulate the nutlets of Amsinckia. There is little in common between the species of the second section of Sonnea and those of the first and typical section, apparently the most important agreement being in the coarse hispid pubescence. Greene

attributed to the second section a fragile caruncular scar similar to that in the typical section, but this attribute I am utterly unable to appreciate. The second section has a peculiar and characteristic nutlet development. The nutlets have a submedial scar that is borne, not at or below the lower end of the ventral keel, but surrounded by and wedged in between the pericarpial margins that form the keel and consequently appearing at first glance as if borne upon it. The striking nutlet difference seems of funamental importance and were other important concomitant characters forthcoming I should feel that the group merits generic recognition. The problem deserves further study and for the present I am following current usage and referring the group to Plagiobothrys where it can form a new section

which may appropriately be called Amsinckiopsis.

The genus Allocarya was erected by Greene, Pittonia i. 10–12, in 1887. In proposing the genus Greene argued that its species "agree admirably in that best mark of a good and natural genus, the habit" and that they possessed "a character very rare in the order, if not indeed unique, that of the lower leaves being not only opposite, but distinctly connate-perfoliate." He states further that "Allocarya is, in truth, much more nearly allied to Plagiobothrys than to Krynitzkia [Cryptantha]. Its nutlets are in general, not very different, being rugose, keeled more or less both dorsally and ventrally, and showing distinct lateral angles." Since its proposal Allocarya has been universally accepted in manuals and floras of western United States. The presence of opposite leaves in its species has been taken as the crucial character. Piper, Contr. U. S. Nat. Herb. xxii. 79 (1920), in his detailed "Study of Allocarya" speaks generally of the genus as follows,—"The genus as delimited by Greene has been generally accepted as valid. It is best distinguished from allied genera by the ventrally keeled nutlets, which are attached basally or suprabasally to a low gynobase, and by having the lowermost leaves opposite." Recently Macbride, Contr. Gray Herb. lix. 34 (1919), wrote, apropos of the reduction of Allocarya to Lappula, as follows:- "Allocarya is most closely related to Plagiobothrys and if Mr. Druce had referred his plant to the latter genus some well-taken arguments for his action could be presented. These genera also, however, are nicely distinct although in nutlet-characters they approach each other closely. The attachment of the nutlets of Plagiobothrys is nearly or quite medial rather than basal or supra-basal and the leaves are never opposite as are the lower ones of Allocarya. The fact that both genera contain numerous species none of which fail in any degree to conform to the generic character in each case is the best argument to my mind as to the validity of those genera." Summing up the above paragraph it may be said that *Allocarya* deserves generic recognition because of its naturalness, this evidenced by its universal acceptance, and because of the constancy and generic value of its habit, the attach-

ment of its nutlets, and its opposite leaves.

Allocarya is said to have a characteristic habit, but at least this is not evident upon a comparison of species so diverse in appearance as A. mollis, A. Scouleri, A. Greenei, A. humistrata, A. californica, and A. stricta. Not only do the species present quite different habits, but they are closely simulated by species commonly referred to Plagiobothrys. For example in gross aspect A. mollis is strikingly like southern forms of P. canescens; A. Greenei and A. Scouleri suggest P. fulvus and P. nothofulvus, while A. scopulorum and A. Cooperi simulate forms of P. californicus. It can be positively said that the species of Allocarya do not have a common and distinctive aspect, and furthermore that they are not as a group habitally distinct from Plagiobothrys.

The attachment of the nutlet is not distinctive of Allocarya. This is at once evident upon a comparison of the fruit of A. Greenei with that of P. fulvus, for in these species there is a remarkable agreement in size and shape of the nutlets and in the arrangement of keels and ridges upon them. Most striking of all is the occurrence in both species of similarly placed, very similar excavated scars. A Patagonian Allocarya described further along in this paper has nutlets closely approximating in both form and attachment those of P. Torreyi. The nutlets of P. Torreyi are also suggested by those of A. mollis. Allocarya does present extremes in fruit developments such as the elongate lance-like basally attached nutlets of A. stipitata and allies, but these are but culminations of tendencies which intermediate developments bring very close to the conditions present in various species of Plagiobothrys.

It would seem that the claims of Allocarya for generic recognition must rest upon the occurrence of opposite leaves on its species. This condition seems to have been at least vaguely realized by the authors who have maintained the genus, for in every keyed or descriptive account of the group the presence of opposite leaves has been given as its crucial character. What has not been realized, however, is that opposite leaves are not particularly characteristic of Allocarya, the development being quite evident and exactly similar in mature plants of Cryptantha rostellata and C. affinis, as well as in the younger stages

of most other species of Cryptantha. Furthermore the character is well developed and very obvious in P. Pringlei and in most (not all) specimens of P. californicus. Allocarya Greenei shows unmistakable relationships with P. fulvus and is probably the nearest relative of that species, vet because it has opposite lower leaves it is forthwith put into another genus. This species alone seems a good argument against the genus Allocarya. It should also be realized that opposite leaves occur in varying abundance in the several species of Allocarya, some having all the leaves opposite, others having a less large proportion or only the lowest pair opposite. In other words there seems to be a complete set of intermediate stages which connect up morphologically the completely opposite-leaved Allocaryas with the completely alternate-leaved Plagiobothrys. In the light of the unquestionably close relation between Allocarya and Plagiobothrys, and the varying proportion of opposite leaves in the species of Allocarya, it is most evident that the use of opposite leaves as a generic criterion is arbitrary and that the currently accepted line of cleavage between the genera is neither a profound nor a particularly natural one. Summing up the present paragraph it can be said that opposite leaves are considered a poor diagnostic character in the present instance because they are present or absent in certain species of Plagiobothrys, and because their absence in Plagiobothrys seems but the culmination (or the beginning) of the variable quantitative development of the character in Allocarya.

Although much stress has been placed on the fact that Allocarya has remained unchallanged since its promulgation over thirty years ago, the fact is significant only as it shows the lack of study of generic relations within the eritrichioid borages. The genus may have had wide acceptance, but it has not had repeated critical reconsiderations. The last critical study of the generic lines of the group was by Greene. Greene's grasp of the relations and characters of the western American eritrichioid borages is undeniable, but unfortunately his subdivisions within the borages, as in many other families, represent intergrading or scarcely distinct groups which seem better treated as sections or

subgenera.

Plagiobothrys can not be separated from Allocarya, Echidiocarya, and Sonnea by developments possessed by the latter genera, and neither can it be distinguished from the enumerated genera by peculiar structures of its own. The character usually given as generically diagnostic for Plagiobothrys is the occurrence of a "caruncular scar" upon its nutlets. However, this character is probably best

developed in Sonnea glomerata and S. hispida, and is present although weakly developed in Allocarya Scouleri and in the Patagonian Allocarya described in this paper. In A. Greenei the character fails completely, for the scar in that species is essentially the same as the scar in P. fulvus. Although it can be said that the caruncular scar reaches a high development in Plagiobothrys, it can not be said that it characterizes the genus or reaches its greatest development there.

Although not previously pointed out, the species of true *Plagio-bothrys* are fairly well characterized by the position of the scar. In *Plagiobothrys* the nutlet-scar is in a broad shallow transverse groove, and distinct from the ventral keel and below the level of it. In *Allocarya* and *Echidiocarya* the scar is usually contiguous with the ventral keel and either flush with it or projected from it on a stipe-like base. As with most other characters its universality is destroyed by *Allocarya Greenei*, that species which is an *Allocarya* only in its lower leaves.

Because of the lack of consistently diagnostic characters which would separate them, Allocarya, Echidiocarya, and Sonnea are all merged with *Plagiobothrys*. The result is a genus which appears to be a very natural one, and one which admits of great precision in definition. The amplified *Plagiobothrys* is at once distinguished from its nearest relatives, Cryptantha and Oreocarya, by the lack of a pronounced longitudinal ventral groove, and the possession instead of a well developed ventral keel and a definitely circumscribed small scar. The gynobase is a pyramid or low frustum and very much shorter than the nutlets, and is not subulate and about equalling the nutlets as in Cryptantha and Oreocarya. The nutlets in Plagiobothrys are commonly keeled and usually rugose, whereas in Cryptantha and Oreocarya they are rarely if ever keeled and the roughenings usually tuberculate or muricate. The pubescence in Plagiobothrys is mostly appressed and is less stiffly spreading than is the hispid indument characteristic of Cruptantha.

In order intelligently to select the specific names which should be transferred to *Plagiobothrys* it has been necessary to go into the intrageneric classification. As a result of this study and with the hope that it will give the paper a wider usefulness I have included a rough key to the accepted species. It is not pretended that the present paper supplies a finished study of the specific classification, but rather only a hastily prepared conservative synopsis which I hope may be preliminary to a future detailed descriptive account. Although it seems certain that future detailed work would cause the

reduction of some of the species here recognized, it is believed that the equally certain recognition of species here reduced will keep the total number of recognized species about as here given. The chief result of a protracted study would be the clarifying of specific lines and the naming and classifying of the abundant intraspecific variation.

During the preparation of this paper I have had Professor Piper's "Study of Allocarya" continually at hand, and have given his suggested classification a careful study. His treatment is based almost entirely upon the shape, sculpturing, and attachment of the nutlets; characters upon which he comments as follows:- "The nutlet characters seem remarkably constant, and in mixed gatherings serve perfectly to segregate the species. Relying on the constancy of the nutlet characters in particular, it is necessary to recognize additional species." The number of recognized species was increased from eighteen to seventy-nine. Although Piper's revision is based upon a wealth of material and shows every evidence of a patient and prolonged investigation I find that I am unable to follow it since I differ from its author in a fundamental point; i.e. I believe that the sculpturing and markings of the nutlets in Allocarya had best be considered excessively variable and consequently of minor if of any importance in the characterization of species. I am led to believe as I do because surprisingly numerous nutlet variations may be discovered in very small areas among plants remarkably uniform in aspect and in all other characters save those of nutlets. Piper has taken the nutlets as constant and has practically disregarded all other characters. whereas I have felt that a paralleling of several vegetative characters were more important than the variation of a single fruiting structure. The nutlet variations of Allocarya have been patiently worked out by Piper and named as species. These minute, very numerous microscopic species appear unpractical and seem justifiable only if they greatly increase the precision in identification and result in a more natural classification. Unfortunately even after considerable familiarity with his key to the numerous species I find it difficult to name plants satisfactorily according to Professor Piper's treatment, and I am compelled to believe that the minute nutlet sculpturings merely appear constant because their great arc of variation has been broken up and specific names associated with very short segments of variation. Though the classification seems carried to an unpractical extreme I would be glad to accept it were it clearly natural. Distribution, I feel, supplies one of the best tests of the naturalness of a species. By the disregard which Piper's species show for the principles underlying Jordan's Law, Science n. s. xxii. 547 (1905), I feel forced to consider them, in a large part, to be unnatural entities. It is highly significant that by largely disregarding nutlet characters and basing species upon obvious external characters that a classification can be made which does not place closely related species together in the same small area. The four species of Allocarya recently published by Brand, Fedde Repert. xviii. 312 (1922), should also make one realize that the acceptance of species based upon unit nutlet characters will undoubtedly lead to the further multiplication of species, for future collecting is bound to turn up unnamed developments and a more careful examination of Piper's specimens is certain to reveal unnamed variations. This will soon make Allocarya one of those genera that are overdivided and hopelessly snarled, and one in which an identification is but rarely attempted—then only with a sigh—and never accomplished with either expedition or with confidence.

Plagiobothrys F. & M. Ind. Sem. Hort. Petrop. ii. 46 (1835). Echidiocarya Gray, Proc. Am. Acad. xi. 89 (1876). Sonnea Greene, Pittonia i. 22 (1887). Allocarya Greene, Pittonia i. 12 (1887).

KEY TO SPECIES.

Leaves all alternate. Caruncle of nutlet elongate, apparently extending along crest of ventral keel; nutlets trigonous, Amsinckia-like. § Amsinckiopsis. Nutlets irregularly rugose; corolla 4–7 mm. broad. Inflorescence elongated, loosely flowered; plant 1–4 Caruncle ovate or orbicular, at or below end of ventral keel. Caruncle weakly developed, borne at tip of a short ventral stipe; nutlets lacking a broad transverse ventral Caruncle well developed, sessile on nutlet, in a broad shallow transverse ventral groove. Inflorescence glomerate; caruncle fragile, at or above the middle of nutlet; basal leaves lacking at maturity of plant. § Sonnea. Nutlets dark, dull, conspicuously rugose and tuberculate, 1–2.3 mm. long. 4. *P. hispidus*. Nutlets light colored, somewhat shiny, nearly smooth, ous, at or below middle of nutlet; basal leaves evident at maturity of plant. § Euplagiobothrys. Calyx circumscissile in fruit, less than 4 mm. long; lobes usually connivent over fruit; usually only 1-2 nutlets developing. Inflorescence a long simple bracteate raceme; stems usually about 2 dm. long, ascending,

hispid, branched mainly below middle; nutlets highly arched in lateral outline, 1-2.5 mm.
long; corolla 3 mm. broad 7. P. arizonicus.
Inflorescence furcate, bracteate only at base if at
all; stems strictly erect, about 3.5 mm. high,
appressed hispid-villous, branched only above
middle if at all; nutlets low and flattened in
lateral outline, 2–3 mm. long; corolla 3–9 mm.
broad
Calyx not circumscissile or if so the strongly ac-
crescent calyx over 4 mm. in length; calyx-lobes
erect or spreading; 4 nutlets usually developing. Nutlets with a conspicuous annular caruncle,
2.3–3.3 mm. long; calyx cleft to near base,
fulvescent; corolla-tube slightly exceeding
calyx 8. P. fulvus.
Nutlets with solid caruncle, less than 2.3 mm.
long; calyx cleft 2/3 to base.
Transverse dorsal crests of nutlets very narrow
and sharp, with medial keel enclosing poly-
gonal granulate areas.
Corolla-tube shorter than calyx; plants erect
to prostrate, comparatively coarse-stem-
med; leaves 3-7 mm. broad; Californian.
Plant dye-stained throughout; nutlets 2 9. P. catalinensis. Plant dye-stained if at all only at base;
nutlets 4
Corolla-tube equalling or slightly exceeding
the calyx; plants very slender, strict or
ascending; leaves 1.5-2.5 mm. broad;
Chilian
Transverse dorsal crests of nutlets very low and
broad, separated only by low lineate
grooves.
Nutlets ovate, usually constricted only at
apex, the base being rounded or rarely weakly constricted, dark-colored; plant
dye-stained
equal constrictions at base and apex,
glassy; plant only exceptionally dye-
stained.
Calyx 5-7 mm. long and nearly as wide;
nutlets 2-2.7 mm. long; inflorescence
bracteate; plant coarse, few-stemmed.13. P. shastensis.
Calyx 3-5(-7) mm. long, and about half as
broad; nutlets 1.5–2 mm. long; slender, many-stemmed plants14. P. tenellus.
Leaves opposite at least below.
Nutlets attached to gynobase by a more or less well de-
veloped stipe-like ventral projection. § Echidiocarya.
Stipe-like base about equaling body of nutlet, frequently
united into pairs
Stipe-like base less than half length of body of nutlet,
never joined into pairs
Nutlets attached directly to gynobase, without a definite
stipe-like ventral prolongation. § Allocarya.

Plant perennial.
Flowers solitary in the axils.
Leaves linear; style greatly surpassing the nutlets. 17. P. Kunthii.
Leaves oblance-linear; style shorter or about equaling
nutlets.
Leaves obtusish usually strigose-hispidulous 10-
15 mm. long; corolla small, 2-3 mm. broad;
compact alpine plants
Leaves acute, somewhat ciliate-strigose along mar-
gins and below on midrib; corolla 4-5 mm.
broad: loosely branched submentance plants 10 D livitaline
broad; loosely branched submontane plants. 19. P. linifolius.
Flowers in axillary glomerules or racemes, frequently
terminal; leaves linear.
Style greatly surpassing the nutlets
Style about equaling nutlets or evidently exceeded
by them.
Pedicels becoming 2-5(-8) mm. long; mature
herbage glabrate.
Leaves 2-2.5 mm. long; stems simple; calyx-
lobes narrowly oblong, acutish, 3 mm. long.
21. P. mexicanus.
Leaves 4.4-5 mm. long; stems branched above;
calyx-lobes narrowly linear, obtusish, 2 mm.
long
Pedicels evidently less than 2 mm. long; mature
herbage somewhat strigose-hispid.
Leaves 25(20)-60 mm. long; fruiting calyx 3
mm. long, lobes very narrowly linear; in-
florescence a rather loose elongated raceme
15-90 mm long sponsols broatest
15-20 mm. long, sparsely bracteate or
naked
long lobes long lines lines in a long long long long long long long long
long, lobes lance-linear; inflorescence a
dense stout glomerule, 5–10 mm. long, leafy
Plant annual bracted throughout
Plant annual.
Spikes geminate
Spikes solitary.
Dorsal keel extending down length of nutlet, usually
well developed.
Scar of nutlet solid, not below level of ventral keel;
nutlets vitreous; Australian 26. P. australasicus.
Scar of nutlet deeply excavated, below level of
ventral keel; nutlets usually dull or glossy:
Californian.
Nutlets 1-1.5 mm. long, weakly keeled dorsally;
stems prostrate, much branched; fruiting
calyces crowded
Nutlets 1.5-2 mm. long, strongly keeled dorsally;
stems erect or ascending, loosely branched;
fruiting calyces remote
Dorsal keel not extending down to the middle of the
nutlet, usually weakly developed.
Scar of nutlet nearly medial; plant dye-stained.
Scar of nutlet at or near the base; plant not dye-
stained.

Ventral keel of nutlet sunken in a longitudinal
groove; pedicels of lower fruiting calyces
5-20 mm. long, slender, spreading or re-
curved.
Nutlets smooth and shiny, ovoid30. P. lithocaryus.
Nutlets roughened, somewhat compressed.
31. P. Chorisianus.
Ventral keel of nutlet prominent, not in a groove; pedicels of lower fruiting calyces
groove; pedicels of lower fruiting calyces
less than 5 mm. long, coarse, stiff, strict or
ascending.
Midrib of calyx-lobes becoming indurated and
enlarged in fruit; calyx sessile or sub-
sessile by a narrowly conic base, usually
asymetrical in fruit.
Plant very stout and fleshy
Plant not stout and fleshy.
Nutlets transversely rugose 33. P. humistratus.
Nutlets reticulately rugose.
Branches strict or ascending34. P. stipitatus.
Branches prostrate or widely spreading.
Branches stiff; bracts few, mainly
near base
Branches merely stiffish; bracts more
or less throughout inflorescence. 36. P. Nelsoni.
Midrib of calyx-lobes scarcely indurate or
thickened in fruit; calyx-lobes tending to
differ sharply from the tube in structure;
calyx usually pedicellate, symetrical,
base broadly conic.
Plant sparsely hispid, lacking appressed
hairs.
Calyx 2.5-4 mm. long, short pedicellate;
calyx-lobes 1–2 mm. long, much sur-
passed by corolla; inflorescence
naked or with 1 or 2 bracts39. P. Parishii.
Calyx 4-6 mm. long, subsessile; calyx-
lobes 2–3 mm. long, about equaling
corolla; inflorescence conspicuously
leafy bracted
Plant with pubescence at least in part of
appressed hairs.
Calyx-lobes spreading or reflexed in fruit,
usually twice length of fruit.
Nutlets reticulately rugose; scar large. 37. P. plebejus.
Nutlets transversely rugose; scar me-
dium size
Calyx-lobes connivent to ascending, 1-2
times length of nutlets.
Calyx firm, strict; plant stiffly erect,
glabrate
Calyx herbaceous, tending to spread;
plant prostrate to erect but not
stiff, usually densely pubescent.
Calyx-lobes 2–3 mm. long, 2 times
length of nutlets

Leaves extremely narrow, 1-1.5 mm. broad; inflorescence prac-Leaves broadly linear, 2-5 mm. broad; inflorescence leafy-

bracted.

Nutlets not at all rugose or

Nutlets rugose or granulate or both......44. P. orthocarpus.

Calyx-lobes 1-2 mm. long, 1-1.5 times length of nutlets.

Leaves fleshy, terete. 45. P. mesembryanthemoides. Leaves herbaceous, flattened.

Nutlets rugose.

Nutlets transversely rugose, rugae usually low and broad; inflorescence usu-

ally naked above....47. P. scopulorum.

Nutlets reticulately rugose, rugae usually narrow and high; inflorescence usually leafy-bracted.

Plant closely prostrate....48. P. Lechleri. Plant erect or ascending. 49. P. procumbens.

1. Plagiobothrys Kingii (Wats.) Gray, Proc. Am. Acad. xx. 281 Eritrichium Kingii Wats. Bot. King Exped. 243, t. 23 (1871). Sonnea Kingii Greene, Pittonia i. 23 (1887). Krynitzkia Kingii Wats. acc. to Hillman, Agric. Exper. Sta. Nev. Bull. xxiv. 71 (1894).—Western Nevada and adjacent California. Apparently a rare species and known only from the vicinity of southern Washoe County, Nevada. It is well marked by its coarse spreading pubescence, large flowers, and naked geminate or ternate racemes.

2. P. Harknessii (Greene) Nels. & Macbr. Bot. Gaz. lxii. 143 (1916). Sonnea Harknessii Greene, Pittonia i. 23 (1887).—Eastern Oregon to Inyo County, California, and eastward to northwestern Utah. Much more common than the last and perhaps only a form

3. P. Jonesii Gray, Synop. Fl. ed. 2, ii. pt. 1, 430 (1886). Sonnea Jonesii Greene, Pittonia i. 23 (1887).—Eastern border of Southern California. Specimens from Inyo County, California, differ from the type in their prostrate habit and may represent an unnamed variety or species. The nutlets of this species remarkably simulate those of Amsinckia tessellata. Because of this Jones, Contr. W. Bot. xii. 57 (1908), wrote that "Plagiobothrys Jonesii Gray is an Amsinckia in every thing but the flowers, which are white. It has the tessellated pavement-like nutlets of A. tessellata and a little sharper

rugae, and illustrates again the very slim foundation on which some Borraginaceous genera rest." These ideas were accepted and repeated by Nelson and Macbride, Bot. Gaz. lxii, 143 (1916), in the following form,-"In this connection Jones has called attention to the fact that P. Jonesii . . . is an Amsinckia in everything but its white flowers. . . . The pubescence of P. Jonesii and the tessellated nutlets surely suggest a relationship to A. tessellata, but the white and short corollas that are so widely at variance with the long yellow ones of Amsinckia are perfectly congeneric with the Sonnea section of Plagiobothrys." The above quotations show a striking lack of acquaintance with the real characters of Plagiobothrys and Amsinckia and are examples of hasty generalizations based upon superficialities. Plagiobothrys Jonesii and immediate relatives simulate species of Amsinckia in the form of nutlets, but are clearly not directly related in that genus, for like other species of Plagiobothrys they have undivided cotyledons and short appendaged corollas.

4. P. HISPIDUS Gray, Proc. Am. Acad. xx. 286 (1885). Sonnea hispida Greene, Pittonia i. 22 (1887).—Eastern Oregon southward through northeastern California and extreme western Nevada to the

region about Mono Lake.

4a. P. hispidus, var. foliaceus (Greene), comb. nov. Sonnea foliacea Greene, Pittonia i. 222 (1888). P. foliaceus Nels. & Macbr. Bot. Gaz. lxii. 143 (1916).—Known only from Washoe Mts., Nevada. Nelson and Macbride speak of the "dorsal depressions of the nutlets" as being "particularly unique" for this plant, but I find that exactly similar developments are frequent in P. hispidus, var. genuinus and in P. glomeratus. The nutlets of the var. foliaceus are 2-2.5 mm. long, instead of 1.5-2 mm. long as in the var. genuinus, and appear to be elongated so that the scar appears decidedly above the middle rather than near the middle of the nutlet. The dorsal surface of the nutlets in the var. foliaceus lack the coarse papillae or irregular short elongate roughenings characteristic of the var. genuinus, instead showing a maximum development of the granulations which in the case of the var. genuinus are crowded by the larger rugosities nearly to extinction. Greene's species is not given specific recognition because it is only known from the type collection. It is possible that the plant is a hybrid between P. hispidus and P. glomeratus.

5. P. GLOMERATUS Gray, Proc. Am. Acad. xx. 286 (1885). Sonnea glomerata Greene, Pittonia i. 22 (1887).—Western Nevada, rare.

Characterized by its large rather smooth nutlets.

6. P. NOTHOFULVUS Gray, Proc. Am. Acad. xx. 285 (1885). Eritrichium nothofulvum Gray, l. c. xvii. 227 (1882).—Frequent over the length of California and locally reaching to the Columbia River Valley.

7. P. ARIZONICUS (Gray) Greene in Gray, Proc. Am. Acad. xx. 284 (1885). Eritrichium canescens, var. arizonicus Gray, l. c. xvii. 227 (1882).—Western New Mexico to Southern California. The species grows in arid situations, and in California is primarily a plant of the desert from which it occasionally extends through the low passes to the dryest of the transmontane valleys. The species is quite variable as to nutlets, and in calyx and habit makes a close approach to, if it does not actually intergrade with P. canescens.

8. P. fulvus (H. & A.), comb. nov. Myosotis fulva H. & A. Bot. Beech. 38 (1830). Eritrichium fulvum A.DC. Prodr. x. 132 (1846). M. alba Colla, Mem. Acad. Torino xxxviii. 128, t. 42 (1835). P. rufescens F. & M. Ind. Sem. Hort. Petrop. ii. 46 (1835 or early 1836). E. asperum Phil. Anal. Univ. Chile xliii. 516 (1873). E. laxiflorum Phil. l. c. xc. 527 (1895). P. rufescens, var. laxiflorus Reiche, Anal. Univ. Chile cxxi. 812 (1908). E. Rengifoanum Phil. l. c. xc. 529 (1895). P. rufescens, var. Renjifoanus Reiche, l. c.—Central Chile.

8a. **P. fulvus**, var. **campestris** (Greene), comb. nov. *P. campestris* Greene, Pittonia ii. 282 (1892). *P. rufescens*, var. *campestris* Jeps. Fl. W. Midd. Calif. 446 (1901). *P. californicus* Greene, Pittonia ii. 231 (1892); not Greene (1887).—California from San Luis Obispo County and the upper San Joaquin Valley northward through the Sacramento Valley to southern Oregon. Through a misunderstanding Gray, Proc. Am. Acad. xx. 282 (1885), cited a specimen of *P. rufescens* as from "near Los Angeles." The Nevin collection upon which this record was based actually came from "n. Sacramento Co." The Oregon collections may represent a distinct variety, their nutlets being dark in color and nearly lacking the transverse dorsal rugae. The Californian plants do not differ in fruit from the Chilian ones, and it is only because the southern plants seem more slender than the northern material that the latter is put into a distinct variety.

9. P. CATALINENSIS (Gray) Macbr. Proc. Am. Acad. li. 546 (1916). P. arizonicus, var. catalinensis Gray, Synop. Fl. ed. 2, ii. pt. 1, 431 (1886).—Endemic on Santa Catalina Island, California. This species is intermediate in its characters between P. canescens and P. arizonicus, having the few nutlets and dye-stained herbage of the latter, and the spreading non-circumscissile calyx of the former. As dye is frequently developed in indubitable P. canescens (cf. Heller

7758 from Mohave, Calif.), and specimens of the var. apertus frequently have imperfectly circumscissile calyces, the claims of *P. arizonicus* and *P. catalinensis* to specific rank are clouded ones, the two species being maintained only for convenience pending further study.

10. P. CANESCENS Benth. Pl. Hartw. 326 (1849). Eritrichium canescens Gray, Proc. Am. Acad. x. 57 (1874). P. microcarpa Greene, Pittonia i. 21 (1887). P. canescens, var. apertus Greene, l. c.—Ranging the length of California. The typical phase of this species is the small, 1.5–2 dm. high, erect-growing plant which is most common in the Sacramento Valley. The var. apertus is best restricted to the large prostrate or subprostrate plant, usually with stellately spreading calyx-lobes, which is most common in the upper San Joaquin Valley. There are numerous other unnamed variations.

11. P. TINCTORIUS (R. & P.) Gray, Proc. Am. Acad. xx. 283 (1885). Lithospermum tinctorium R. & P. Fl. Peruv. ii. 4, t. 114 (1799). Eritrichium tinctorium A.DC. Prodr. x. 132 (1846). L. myosotoides Lehm. Asperif. ii. 319 (1818). L. tingens R. & S. Syst. iv. 44 (1819). E. verrucosum Phil. Linnaea xxix. 17 (1857).—Central Chile.

12. P. Torreyi Gray, Proc. Am. Acad. xx. 284 (1885). Eritrichium Torreyi Gray, l. c. x. 58 (1875). Cryptanthe Torreyi Rydb. Mem. N. Y. Bot. Gard. i. 331 (1900).—California; apparently confined to vicinity of Yosemite Valley.

12a. P. Torreyi, var. diffusus, var. nov., laxus prostratus; caulibus ad basin florigeris; floribus numerosis; calyce fructifero 3-4 mm. longo 3-4.5 mm. lato; bracteis grandis conspicuissimis.—Cali-FORNIA: Sierra Valley, Lemmon. Donner Lake, Heller 6986 (TYPE, Gray Herb.). Sunnyside, Lake Tahoe Region, Eastwood 1067. About Tallac, July 1904, M. S. Baker. Echo Camp on Lincoln Highway, Heller 12,158. Yosemite, 1878, Lemmon. Upper San Joaquin, Madera Co., 1895, Congdon. Okenden, Pine Ridge, Fresno Co., Hall & Chandler 273. Volcano Cr., Tulare Co., Hall & Babcock 5317. This is the most common and widely distributed phase of P. Torreyi. It was mentioned by Gray when he described the species, but was not named by him. The type of P. Torreyi is evidently Torrey 338 and that is the slender erect-growing plant with few leaves and bracts which is frequently collected in the region about Yosemite The typical phase of P. Torreyi and the var. diffusus are quite dissimilar in aspect, differing as they do in direction of growth. leafiness, amount of branching, and size of bracts. Young forms of var. diffusus might be confused with typical Torreyi, but may be usually distinguished by having the lower floral bracts three or more times as long as the calyx.

altior apicem versus laxe ramosus; bracteis inconspicuis paucis; nuculis ovatis basin versus paulo constrictis; habitu *P. tenelli* sed caulibus colorantibus.—California: Greenhorn Pass, alt. 4–5000 ft., *Purpus 5542* (Type, Univ. Calif.). Greenhorn Range, Kern Co., alt. 5000 ft., *Hall & Babcock 5041*. This variety has a characteristic habit, but it is primarily distinguished by its nutlets which, while most like those of *P. Torreyi*, have weakly constricted bases and so suggest the nutlets of *P. tenellus*. The new variety can be looked upon as a connecting link between *P. tenellus* and *P. Torreyi*, but due to its combining of characters it suggests a hybrid and so, pending further information, I am leaving the status of its possible parents unchanged.

13. P. SHASTENSIS Greene in Gray, Proc. Am. Acad. xx. 284 (1885).

—California from the lower San Joaquin Valley northward to southern Oregon; not common. A very near relative of *P. tenellus* and perhaps only a rankly growing strain of it, but usually to be recognized by its large fruiting calyces and nutlets, erect simple few-flowered stems, and soft pubescence. It superficially suggests *P. canescens* but may

be readily told by its very different nutlets.

14. P. TENELLUS (Nutt.) Gray, Proc. Am. Acad. xx. 283 (1885). Myosotis tenella Nutt. in Hook. Kew Jour. Bot. iii. 295 (1851), nom. subnudum. Eritrichium tenellum Gray, l. c. x. 57 (1875). P. echinatus Greene, Pittonia iii. 262 (1898). P. asper Greene, l. c. P. humifusa Jones, Contr. W. Bot. xiii. 7 (1910).—British Columbia to northern Utah and Nevada, and southward through the coastal drainage of California to northern Lower California. This is the most widely ranging and most variable of the species of true Plagiobothrys. Greene has named a number of forms, but these do not seem striking or constant enough to warrant their recognition. Plagiobothrys asper is a conspicuously hispid phase, which is not geographically correlated and intergrades so gradually and completely with the normal villous forms that its recognition even as a forma seems unpractical. Plagiobothrys humifusus is a compact form of P. asper.

14a. **P. tenellus**, var. **parvulus** (Greene), comb. nov. *P. parvulus* Greene, Pittonia iii. 261 (1898).—California; along the South Coast Ranges from San Francisco Bay Region to San Luis Obispo County. This is the prevailing form of the species in the area of its occurrence, and from its geographic correlation perhaps deserves minor recognition. It differs from other forms of the species by having its nutlets 1-1.5 mm. long and its fruiting calvees 2 mm. long or less.

In the typical form of the species the nutlets are 1.5-2 mm. long

and the calvees measure 2-3 mm. in length.

14b. P. tenellus, var. colorans, comb. nov. P. colorans Greene Pittonia iii. 262 (1898).-Known only from extreme northern California. A poorly understood plant which appears to be only a dyestained form of P. tenellus.

15. P. Pringlei Greene, Pittonia i. 21 (1887). Echidiocarya arizonica Gray, Proc. Am. Acad. xi. 89 (1875).—Southern Arizona

and adjacent Sonora.

16. P. CALIFORNICUS (Gray) Greene, Bull. Calif. Acad. ii. 407 (1887). Echidiocarya californica Gray, Proc. Am. Acad. xii. 164 (1877). P. Cooperi Gray, l. c. xx. 285 (1885).—Southern California and northern Lower California. Differing from the last only in the length of the stipe-like base of the nutlet. There are the following well marked varieties.

Corolla 4-7 mm. broad; pubescence fine, appressed, usually

Leaves narrowly linear, 2-2.5 mm. broad; pubescence fine,

canescent......var. gracilis.

Leaves oblanceolate, 3–5 mm. broad; pubescence coarse,

Racemes dense, hidden among the leaves var. ursinus. Racemes elongated, projected from among the leaves and evident.....var. fulvescens.

- 16a. P. californicus, var. genuinus, var. nov.—California: Near San Gabriel, Brewer 147. Near Upland, Johnston 1839. San Bernardino, Parry 213. Corona, Johnston 1876. Las Flores, Abrams 3276. La Jolla, Clements 110. San Diego, Spencer 126; Brandegee 1637. LOWER CALIFORNIA: Tia Juana Valley, April 1882, Pringle. San Rafael Valley, April 1885, Orcutt. The above cited suite of selected specimens covers the range of genuinus. It is the common form on the grassy hillsides on the coastal drainage of Southern California. The stems are long, lax, and strigose-canescent. It has conspicuous corollas, large fruiting calyces, and broad oblanceolate leaves.
- 16b. P. californicus, var. gracilis, var. nov., hispidulosus minutiflorus; caulibus pergracilibus; foliis lanceolato-linearibus acutis sparsis; sepalis angustis.—California: La Jolla, Clements 111. San Diego, Brandegee 1658 (TYPE, Gray Herb.); Orcutt 1014, in part. LOWER CALIFORNIA: Cedros Island, Palmer 711. Without locality Parry. A very well marked variety which apparently grows with var. genuinus and seems much less common. It is perhaps specifically distinct.

- 16c. P. californicus, var. ursinus (Gray), comb. nov. Echidiocarya ursina Gray, Proc. Am. Acad. xix. 90. (1883). P. ursinus Gray, l. c. xx. 285 (1885).—California: San Bernardino Mts., Parish 927 (Type); Munz 5725. San Jacinto Mts., Spencer 1656; Munz & Johnston 5416. Lower California: Without locality. Orcutt 908. A variety inhabiting warm montane valleys and differing from the following only in its congested inflorescence.
- 16d. P. californicus, var. fulvescens, var. nov., hispidus minutiflorus; caulibus elongatis prostratis; foliis oblanceolatis; inflorescentia elongata remotiflora a foliis non obscurata.—California: Santa Barbara, 1888, T. S. Brandegee (TYPE, Gray Herb.). Witch Creek, Alderson 7625. Lower California: Hansen's Ranch, April 1885, Orcutt.
- 17. P. Kunthii (Walp.), comb. nov. Anchusa Kunthii Walp. Nov. Act. Nat. Cur. xix. suppl. 1, 372 (1843). Allocarya linifolia, var. Kunthii Macbr. Proc. Am. Acad. li. 545 (1916). Antiphytum Walpersii A.DC. Prodr. x. 122 (1846). Eritrichium Walpersii Wedd. Chlor. And. ii. 90 (1859).—I associate with this name two Bolivian collections (Mandon 382, 383) in the herbarium of the New York Botanical Garden. The flowers are solitary in the axils and the corollas have very elongate tubes and conspicuous (ca. 4 mm. broad) limbs. The style surpasses the mature nutlets by nearly 2 mm. The pedicels are very slender becoming nearly 5 mm. long. Walpers's description is extremely short and vague, the Mandon collections being associated with it only because they come from Lake Titicaca, the type region of Anchusa Kunthii, and because they alone among the available Peruvian and Bolivian material agree with the "floribus solitariis, axillaribus" of the original diagnosis.
- 18. P. pygmaeus (HBK.), comb. nov. Anchusa pygmaea HBK. Nov. Gen. et Sp. iii. 92 (1818). Eritrichium pygmaeum Wedd. Chlor. And. ii. 89 (1859). Lithospermum alpinum R. & S. Syst. iv. 742 (1819).—Ecuador.
- 19. P. linifolius (Lehm.), comb. nov. Anchusa linifolia Lehm. Asperif. i. 215 (1818). Antiphytum linifolium A.DC. Prodr. x. 121 (1846). Eritrichium linifolium Wedd. Chlor. And. ii. 89 (1859). Krynitzkia linifolia Gray, Proc. Am. Acad. xx. 266 (1885). Allocarya linifolia Macbr. Proc. Am. Acad. li. 545 (1916). Anchusa oppositifolia HBK. Nov. Gen et Sp. iii. 91, t. 200 (1818).—Ecuador.
- 20. P. mollis (Gray), comb. nov. Eritrichium molle Gray, Proc. Am. Acad. xix. 89 (1883). Allocarya mollis Greene, Pittonia i. 20 (1887).—California, along the northern Sierras, and in adjacent Nevada and Oregon.

20a. **P. mollis** var. **vestita** (Greene), comb. nov. Allocarya vestita Greene, Erythea iii. 125 (1895). A. mollis, var. vestita Jepson, Fl. Midd. W. Calif. 442 (1901).—Middle California; rare. Known only from Marin and Tulare counties. It is apparently a geographical variety differing from the typical form of the species in its darker and more loosely reticulate-rugose nutlets.

21. P. mexicanus (Macbr.), comb. nov. Allocarya mexicana Macbr. Contr. Grav Herb. lix. 34 (1919).—Mexico. Known only from the

type collection made in the state of Mexico.

22. P. pedicellaris (Phil.), comb. nov. Eritrichium pedicellare Phil. Anal. Univ. Chile xc. 549 (1895). Allocarya pedicellaris Reiche, Anal. Univ. Chile cxxi. 809 (1907).—Chile. Known to me only by

descriptions.

23. P. humilis (R. & P.), comb. nov. Myosotis humilis R. & P. Fl. Peruv. ii. 5 (1799). Eritrichium humile A.DC. Prodr. x. 133 (1846). Allocarya humilis Greene, Pittonia i. 17 (1887). Amsinckia humifusa Walp. Nov. Act. Nat. Cur. xix. suppl. 1, 371 (1843). Benthamia humifusa Druce, Rep. Bot. Exch. Cl. Brit. Isl. iv. 298 (1916). E. Germaini Phil. Anal. Univ. Chile xc. 550 (1895). Allocarya Germaini Reiche, Anal. Univ. Chile cxxi. 809 (1907).—Peru and Bolivia. Apparently also in Chile, Philippi's species being doubtfully associated with the Peruvian one.

24. P. congestus (Wedd.), comb. nov. Eritrichium humile, var. congestum Wedd. Chlor. And. ii. 88 (1859).—Peru and Bolivia, ap-

parently at high altitudes.

25. P. Scouleri (H. & A.), comb. nov. Myosotis Scouleri H. & A. Bot. Beech. 370 (1840), nom. subnudum. Eritrichium Scouleri A.DC. Prodr. x. 130 (1846). Krynitzkia Scouleri Gray, Proc. Am. Acad. xx. 267 (1885). Allocarya Scouleri Greene, Pittonia i. 18 (1887). E. sessiliflorum A.DC. l. c. 133. A. sessilifolia Greene, l. c. 17. A. hirta Greene, l. c. 161 (1888). A. Scouleri, var. hirta Nels & Macbr. Bot. Gaz. lxi. 36 (1916). A. calycosa Piper, Contr. U. S. Nat. Herb. xxii. 101 (1920). A. figurata Piper, l. c. 101. A. dichotoma Brand in Fedde, Repert. xviii. 313 (1922).—Oregon to southern British Columbia, and apparently also in central Chile.

26. P. australasicus (A.DC.), comb. nov. Eritrichium australasicum A.DC. Prodr. x. 134 (1846). Allocarya australasica Greene, Erythea iii. 57 (1895).—Australia. This is the only extra-American

species of the genus.

27. P. Piperi, nom. nov. Allocarya microcarpa Piper, Contr. U. S. Nat. Herb. xxii. 91 (1920); not P. microcarpus Greene (1887).—

Known only from Mariposa County, California. Superficially nearly indistinguishable from *P. scopulorum*, but in fruit-characters clearly allied with *P. Greenei*.

- 28. P. Greenei (Gray), comb. nov. Echinospermum Greenei Gray. Proc. Am. Acad. xii. 163 (1877). Allocarya Greenei Greene, Bot. San Francisco 259 (1894). A. Echinoglochin Greene, Pittonia i. 15 (1887). A. Austinae Greene, Pittonia i. 18 (1887). A. hystricula Piper, Contr. U. S. Nat. Herb. xxii. 87 (1920). A. acanthocarpa Piper, l. c. 87. A. oligochaeta Piper, l. c. 88. A. echinacea Piper, l. c. 88. cristata Piper, l. c. 89. A. Eastwoodae Piper, l. c. 89. A. glyptocarpa Piper, l. c. 90. A. spiculifera Piper, l. c. 90. A. anaglyptica Piper, l. c. 90. A. papillata Piper, l. c. 91. A. distantiflora Piper, l. c. 91. Occuring over the length of California. Very conspicuously variable in the sculpturing and arming of the nutlets, even in a single locality, and apparently showing no tendency to break up into definite geographic variants. Some of the conspicuous extremes might well be treated as formae. Allocarya glyptocarpa probably is specifically distinct differing from the great mass of P. Greenei in its large corollas and elongate nutlets.
- 29. P. patagonicus, sp. nov., annuus tinctus; caulibus prostratis diffuse ramosis ca. 1 dm. longis sparse breveque villosis; foliis ovatooblongis vel oblongis 13-15 mm. longis 3-5 mm. latis, infimis oppositis; racemis elongatis maturitate remote florentibus cum bracteis foliaceis; floribus albis ca. 2.5 mm. longis; corollae tubo calyce longiori cylindrato, lobis ascendentibus; calyce strigoso-hispido ad basin partito ad anthesin ca. 2 mm. longo fructifero aperto 3-4 mm. longo; nuculis 4 late ovatis ca. 1.5 mm. longis dorso congeste humileque rugosis ventrale carinatis in media parte ad gynobasin humilem adfixis.-ARGENTINA: Patagonia, 50° 3' Lat., 1882, Moreno & Tonini 530 (TYPE, N. Y. Bot. Gard.). San Carlos de Bariloche, 800 m. alt., Buchtien 118 (U. S.). In gross aspect much resembling P. Torreyi, var. diffusus, and in most parts suggesting a true Plagiobothrys rather than a species of Allocarya which it must be because of its opposite lower leaves. It is probably the "Plagiobothrys decumbens" of Macloskie, Fl. Patag. 679 (1905), and perhaps also the "Cryptanthe globulifera" of Skottsberg, Svenska Vet. Akad. Handl. lvi. no. 5, 290 (1916).
- 30. P. lithocaryus (Greene), comb. nov. Krynitzkia lithocarya Greene in Gray, Proc. Am. Acad. xx. 265 (1885). Allocarya lithocarya Greene, Pittonia i. 12 (1887).—California, along the North Coast Ranges. Rare.

- 31. P. Chorisianus (Cham.), comb. nov. Myosotis Chorisiana Cham. Linnaea iv. 444 (1829). Eritrichium Chorisianum A.DC. Prodr. x. 130 (1846). Krynitzkia Chorisiana Gray, Proc. Am. Acad. xx. 267 (1885). Allocarya Chorisiana Greene, Pittonia, i. 13 (1887). E. connatifolium Kell. Proc. Calif. Acad. ii. 163, f. 51 (1862). A. Hickmanii Greene, Pittonia i. 13 (1887). A. myriantha Greene, Erythea iii. 125 (1895). A. Jonesii Brand in Fedde, Repert. xviii. 313 (1922).—California, from San Francisco to Santa Barbara counties.
- 32. P. glaber (Gray), comb. nov. Lithospermum glabrum Gray, Proc. Am. Acad. xvii. 227 (1882). Allocarya glabra Macbr. Proc. Am. Acad. li. 543 (1916). A. salina Jepson, Fl. W. Midd. Calif. 442 (1901).—Middle California, and doubtfully also Arizona. Mrs. Brandegee, Zoe v. 94 (1901), doubts the Arizonian origin of the type of L. glabrum. No undoubted material is at hand from Arizona, and Professor J. J. Thornber of the University of Arizona writes me that he has neither collected such a plant in Arizona nor knows of anyone else having done so. It is possible that the following four species had best be treated as varieties of the present one.

33. P. humistratus (Greene), comb. nov. Allocarya humistrata Greene, Pittonia i. 16 (1887). A. scripta Greene, l. c. 142. A. limicola Piper, Contr. U. S. Nat. Herb. xxii. 97 (1920). A. sigillata Piper, l. c.—Middle California.

34. P. stipitatus (Greene), comb. nov. Allocarya stipitata Greene, Pittonia i. 19 (1887).—Lappula stipitata Druce, Rep. Bot. Exch. Cl. Brit. Isl. v. 38 (1918). A. stipitata, subsp. micrantha Piper, Contr. U. S. Nat. Herb. xxii. 94 (1920). A. ambigens Piper, l. c. 96.—Middle California. There are two conspicuous extremes in flower-size.

35. P. divergens (Piper), comb. nov. Allocarya divergens Piper, Contr. U. S. Nat. Herb. xxii. 92 (1920). A. charaxata Piper, l. c. 96.—

California, from Tulare County to San Diego County.

36. P. Nelsonii (Greene), comb. nov. Allocarya Nelsonii Greene, Erythea iii. 48 (1895). A. leptoclada Greene, Pittonia iii. 109 (1896). A. oricola Piper, Contr. U. S. Nat. Herb. xxii. 92 (1920). A. asperula Piper, l. c. 93. A. Wilcoxii Piper, l. c. 93. A. setulosa Piper, l. c. 93. A. Leibergii Piper, l. c. 95. A. tuberculata Piper, l. c. 95. A. fragilis Brand in Fedde, Repert. xviii. 312 (1922).—Eastern Oregon and northern Nevada and northwestward to Montana and adjacent Saskatchewan.

37. P. plebejus (Cham.), comb. nov. Lithospermum plebejum Cham. Linnaea iv. 446 (1829). Eritrichium plebeium A. DC. Prodr.

x. 133 (1846). Krynitzkia plebeia Gray, Proc. Am. Acad. xx. 266 (1885). Allocarya plebeia Greene, Pittonia i. 16 (1887).—Alaska.

- 38. P. trachycarpus (Gray), comb. nov. Krynitzkia trachycarpa Gray, Proc. Am. Acad. xx. 266 (1885). Allocarya trachycarpa Greene, Pittonia i. 14 (1887). Myosotis californica F. & M. Ind. Sem. Hort. Petrop. ii. 42 (1835). Eritrichium californicum A.DC Prodr. x. 130 (1846). K. californica Gray, l. c. A. californica Greene, l. c. 20; not P. californicus Greene (1887). A. diffusa Greene, l. c. 14. A. interrasilis Piper, Contr. U. S. Nat. Herb. xxii. 108 (1920). A. commixta Brand in Fedde, Repert. xviii. 312 (1922).—Coast Ranges of middle California. The type of K. trachycarpa, Brewer 1007 from Sonoma County, is a good match for authentic specimens of M. californica, and appears to represent the very slender plant with long lax leafy stems and linear spreading calvx-lobes which seems to replace P. scopulorum in the region along the middle Coast Ranges of California. As I have taken it P. trachycarpus may consist of two things, the southern plants seeming to be less diffuse and to have shorter calvx-lobes.
- 39. **P. Parishii**, nom. nov. Eritrichium Cooperi Gray, Proc. Am. Acad. xix. 89 (1883). Krynitzkia Cooperi Gray, l. c. xx. 267 (1885). Allocarya Cooperi Greene, Pittonia i. 19 (1887); not P. Cooperi Gray (1885).—Mohave Desert of California.

40. **P. salsus** (Brandg.), comb. nov. Allocarya salsa Brandg. Bot. Gaz. xxvii. 452 (1899). A. jacunda Piper, Bull. Torr. Cl. xxix. 643 (1902). A. Cusickii, var. jacunda Nels. & Macbr. Bot. Gaz. lxi. 36 (1916).—Nevada and eastern Oregon.

41. P. strictus (Greene), comb. nov. Allocarya stricta Greene, Pittonia ii. 231 (1892).—Northern California, perhaps best restricted to the Calistoga plant.

- 42. P. tenuifolius (Gray), comb. nov. Krynitzkia tenuifolia Gray, Proc. Am. Acad. xx. 267 (1885). Eritrichium tenuifolium Phil. Anal. Univ. Chile xlii. 518 (1873), nom. nudum, & xc. 546 (1895). Allocarya tenuifolia Greene, Erythea iii. 57 (1895). E. humile, var. capitatum Clos in Gay, Fl. Chile iv. 471 (1849). E. tenuifolium, var. longipes Phil. l. c. xlii. 518 (1873). A. tenuifolia, var. longipes Reiche, Anal. Univ. Chile cxxi. 806 (1907).—Chile.
- 43. P. nitens (Greene), comb. nov. Allocarya nitens Greene, Pittonia iii. 108 (1896).—Nevada and Utah, apparently rare. Perhaps only a phase of the next.
- 44. **P. orthocarpus** (Greene), comb. nov. *Allocarya orthocarpa* Greene, Pittonia iv. 235 (1901).—Washington and Nevada, eastward to Utah and Colorado. A rare and poorly understood species.

45. P. mesembryanthemoides (Speg.), comb. nov. Eritrichium mesembryanthemoides Speg. Anal. Soc. Cientf. Argent. liii. 136 (1902). —Patagonia.

46. P. muricatus (R. & P.), comb. nov. Lithospermum muricatum R. & P. Fl. Peruv. ii. 4 (1799). Eritrichium muricatum A.DC. Prodr. x. 132 (1846). Allocarya muricata Reiche, Anal. Univ. Chile cxxi. 810 (1907).—Chile. This plant may be a Cryptantha, although Philippi's note, Anal. Univ. Chile xc. 540 (1895), makes it seem improbable. It is possible that the species should be amplified to include the concepts here called P. procumbens and P. scopulorum.

47. P. scopulorum (Greene), comb. nov. Allocarya scopulorum Greene, Pittonia i. 16 (1887). Eritrichium californicum, var. subglochidiatum Gray in Wats. Bot. Calif. i. 526 (1876). Krynitzkia californica, var. subglochidiata Gray, Proc. Am. Acad. xx. 266 (1885). A. subglochidiata Piper, Contr. U. S. Nat. Herb. xi. 485 (1906). A. hispidula Greene, l. c. 17. A. Cusickii Greene, l. c. 17. A. penicillata Greene, l. c. 18. A. tenera Greene, l. c. iii. 109 (1896). A. cognata Greene, l. c. iv. 235 (1901). A. bracteata Howell, Fl. N. W. Amer. 481 (1901). A. cryocarpa Piper, Contr. U. S. Nat. Herb. xxii. 98 (1920). A. gracilis Piper, l. c. 98. A. laxa Piper, l. c. 98. A. pratensis Piper, I. c. 99. A. cervina Piper, I. c. 100. A. ramosa Piper, l. c. 100. A. vallata Piper, l. c. 101. A. undulata Piper, l. c. 104. A. minuta Piper, l. c. 104. A. scalpta Piper, l. c. 104. A. reticulata Piper, l. c. 105. A. areolata Piper, l. c. 105. A. inornata Piper, l. c. 106. A. media Piper, l. c. 107. A. divaricata Piper, l. c. 107. A. insculpta Piper, l. c. 109. A. dispar Piper, l. c. 109. A. granulata Piper, l. c. 109. A. conjuncta Piper, l. c. 109. A. corrugata Piper, l. c. 110. A. scalpocarpa Piper, l. c. 111.—Western United States and adjacent Canada. This is the most common and widely distributed Allocarya, and that which has mostly borne the name A. californica. It varies considerably in the marking and sculpturing of the nutlets, and to a less extent in habit as well. A careful study will probably cause the recognition of a number of forms here submerged. There is a largeflowered plant on Vancouver Island (e.g. Macoun 56, 680), and a stiffish strict one of western Oregon (e.g. Sheldon 10,577) which may be distinct. Some plants from the Argentine, for the present referred to P. procumbens, seem indistinguishable from certain of the North American specimens.

48. P. Lechleri, nom. nov. Eritrichium albiflorum Griseb. Abhandl. Ges. Wiss. Gött. vi. 131 (1854); not Myosotis albiflora B. & S. in Hook. f. Fl. Antarct. ii. 329 (1847).—Patagonia and Fuego. This

is the plant which has been variously identified as Eritrichium albiflorum (Grisebach, l. c.), E. diffusum (Dusén, Svenska Exped. Magell.
iii. 132 (1900)), and Allocarya procumbens (Skottsberg, Svenska Vet.
Akad. Handl. lvi. 289 (1916)). It was a specimen of this species,
incorrectly identified as Myosotis albiflora, that gave Greene, Erythea
iii. 57 (1895), his reasons for proposing the combination, Allocarya albiflora. Regarding the identity of Myosotis albiflora B. & S. see the

lengthy note by Skottsberg, l. c. 290-291, t. 23, f. 8a-d.

49. P. PROCUMBENS (Colla) Gray, Proc. Am. Acad. xx. 283 (1885). Myosotis procumbens Colla, Mem. Acad. Torino xxxviii. 130 (1834). Eritrichium procumbens A.DC. Prodr. x. 133 (1846). Allocarya procumbens Greene, Pittonia i. 17 (1887). E. tenuicaule Phil. Linnaea xxix. 18 (1857). A. tenuicaulis Macbr. Proc. Am. Acad. li. 544 (1916). E. uliginosum Phil. Anal. Univ. Chile xliii. 519 (1873). A. uliginosa Greene, l. c. 14. E. calandrinioides Phil. Anal. Univ. Chile xc. 541. (1895). E. oppositifolium Phil. l. c. 542. A. oppositifolia Reiche, Anal. Univ. Chile cxxi. 807 (1907). E. polycaule Phil. l. c. 542. E. delicatulum Phil. l. c. 544. E. flavicans Phil. l. c. 544. E. pulchellum Phil. 1. c. 545. E. cinereum Phil. 1. c. 545. A. cinerea Reiche, l. c. 808. E. limonium Phil. l. c. 546. E. graminifolium Phil. l. c. 547. E. illapelinum Phil. l. c. 548. E. bracteatum Phil. l. c. 548. E. vernum Phil. l. c. 550.—Chile and Argentine. This appears to be the South American homologue of P. scopulorum, and like it is very variable in its structures.

III. DIAGNOSES AND NOTES RELATING TO THE SPERMATOPHYTES CHIEFLY OF NORTH AMERICA.

By I. M. Johnston.

The subjoined paragraphs bring together miscellaneous data which have accumulated during the past few months as a by-product of general herbarium work. Considerable time has been spent by the author in ordering up the *Euphorbiaceae* of the Gray Herbarium. As a result of this work it has been found desirable to place on record certain undescribed species which have been detected and to make some new combinations which were needed in order that the naming of the collection could be strictly in accord with the International Rules of Nomenclature. Some time has also been spent in an



Johnston, I. M. 1923. "Studies in the Boraginaceae." *Contributions from the Gray Herbarium of Harvard University* (68), 43–80. https://doi.org/10.5962/p.336051.

View This Item Online: https://www.biodiversitylibrary.org/item/123746

DOI: https://doi.org/10.5962/p.336051

Permalink: https://www.biodiversitylibrary.org/partpdf/336051

Holding Institution

Missouri Botanical Garden, Peter H. Raven Library

Sponsored by

Missouri Botanical Garden

Copyright & Reuse

Copyright Status: Permission to digitize granted by rights holder License: http://creativecommons.org/licenses/by-nc-sa/3.0/
Rights: https://www.biodiversitylibrary.org/permissions

This document was created from content at the **Biodiversity Heritage Library**, the world's largest open access digital library for biodiversity literature and archives. Visit BHL at https://www.biodiversitylibrary.org.